Podarcis lilfordi genome annotation report

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Methods

Genome annotation

Repeats present in the rPodLil1.1 genome assembly were annotated with RepeatMasker v4-1-2 (http://www.repeatmasker.org) using the custom repeat library available for podarcis. Moreover, a new repeat library specific for our assembly was made with RepeatModeler v1.0.11. After excluding those repeats that were part of repetitive protein families (performing a BLAST (1) search against Uniprot) from the resulting library, RepeatMasker was run again with this new library in order to annotate the specific repeats.

The gene annotation of the Lilford's wall lizard genome assembly was obtained by combining transcript alignments, protein alignments and *ab initio* gene predictions. A flowchart of the annotation process is shown in **Figure ANN1**.

Firstly, RNA from five different tissues (heart, kidney, liver, lungs and tail) was obtained and sequenced with both Illumina RNAseq and PacBio IsoSeq. After sequencing, the long and short reads were aligned to the genome using, respectively, STAR (2) v-2.7.2a and MINIMAP2 (3) v2.14 with the splice option. Transcript models were subsequently generated using Stringtie (4) v2.1.4 on each BAM file and then all the models produced were combined using TACO (5) v0.6.3. High-quality junctions to be used during the annotation process were obtained by running Portcullis (6) v1.2.0 after mapping with STAR and MINIMAP2. Finally, PASA assemblies were produced with PASA (7) v2.4.1. The *TransDecoder* program, which is part of the PASA package, was run on the PASA assemblies to detect coding regions in the transcripts. Secondly, the complete proteomes of *Podarcis muralis, Pogona vitticeps* and *Pantherophis guttatus* were downloaded from Uniprot in April 2002 and aligned to the genome using Spaln (8) v2.4.03. *Ab initio* gene predictions were performed on the repeat-masked rPodLil1.1 assembly with three different programs: GeneID (9) v1.4, Augustus (10) v3.3.4 and Genemark-ES (11) v2.3e with and without incorporating evidence from the RNAseq data. The gene predictors were run with trained parameters for human except Genemark, which runs in a self-trained mode. Finally, all the data were combined into consensus CDS models using EvidenceModeler-1.1.1 (EVM) (7). Additionally, UTRs and alternative splicing forms were annotated via two rounds of PASA annotation updates. Functional annotation was performed on the annotated proteins with Blast2go (12). First, a Diamond Blastp (13) search was made against the nr database (last accessed May 2022). Furthermore, Interproscan (14) was run to detect protein domains on the annotated proteins. All these data were combined by Blast2go, which produced the final functional annotation results.

The annotation of ncRNAs was obtained by running the following steps. First, the program cmsearch (15) v1.1 that is part of the Infernal (16) package was run against the RFAM database of RNA families (16) v12.0. Additionally, tRNAscan-SE (17) v2.08 was run in order to detect the tranfer RNA genes present in the genome assembly. Identification of lncRNAs was done by first filtering the set of PASA-assemblies that had not been included in the annotation of protein-

coding genes to retain those longer than 200bp and not covered more than 80% by a small ncRNA. The resulting transcripts were clustered into genes using shared splice sites or significant sequence overlap as criteria for designation as the same gene.

Results

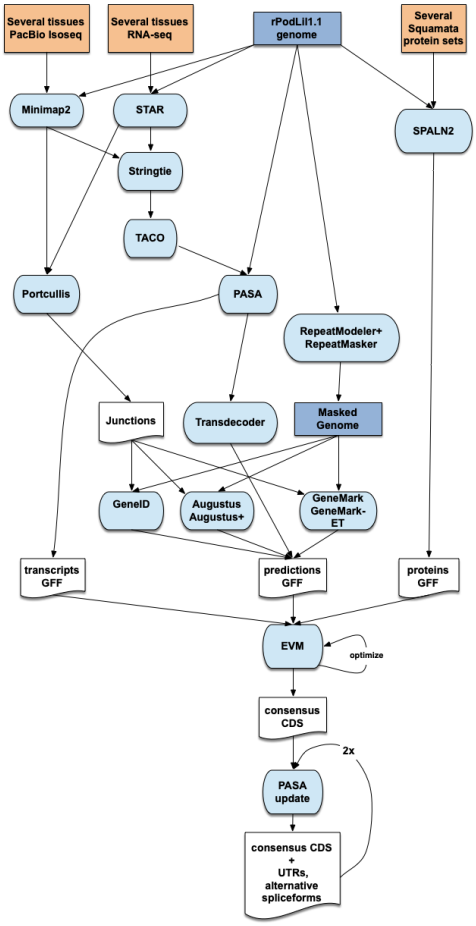
Genome annotation

In total, we annotated 25,678 protein-coding genes that produce 43,594 transcripts (1.7 transcripts per gene) and encode for 38,631 unique protein products. We were able to assign functional labels to 72% of the annotated proteins. The annotated transcripts contain 10.9 exons on average, with 91% of them being multi-exonic (**Table ANN1**). In addition, 47,087 non-coding transcripts were annotated, of which 12,794 and 34,293 are long and short non-coding RNA genes, respectively.

Table ANN1: Genome annotation statistics

	PODLIA annotation
Number of protein-coding genes	25,678
Median gene length (bp)	13,439
Number of transcripts	43,594
Number of exons	247,241
Number of coding exons	232,510
Median UTR length (bp)	2,138
Median intron length (bp)	1,296
Exons/transcript	10.9
Transcripts/gene	1.7
Multi-exonic transcripts	91%
Gene density (gene/Mb)	17.58

Figure ANN1: workflow of the genome annotation process



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